

Patient Information**1. PATIENT IDENTIFICATION**

Hospital:

UPN:

Usual residence:

Postcode:

Race:

Indigenous status:

Consented: Y | N

Name id (optional):

DOB: __/__/__

Sex:

CIBMTR ID (CRID):

AID:

Cell Therapy Pre-Infusion**1. PATIENT IDENTIFICATION**

Referral centre:

Referring doctor:

Date of first referral for cell therapy: __/__/__

2. CELL THERAPY

Participating in CT clinical trial: Y | N

If yes: Corporate Investigator initiated other _____

Study id number:

Complete copies of above questions if on multiple trials

If no, reason why not in clinical trial:

 Institutional guidelines Hospital exemption Compassionate useProduct funding: Clinical Trial MBS MTOP Self-funded**3. PRIOR CELL THERAPY (CT)**

This is first course of cell therapy (non HCT): Y | N | Unk

If no: reported to: ABMTRR CIBMTR EBMT

Number prior CTs: ____ Date of CT: __/__/__

Where performed: _____ Indication: _____

Cell source(s): Auto Allo-unrelated Allo-related*Complete copies of above questions if >1 prior CT***4. PRIOR TRANSPLANT (HCT)**

Received prior HCT: Y | N | Unk

If yes, reported to: ABMTRR CIBMTR EBMT

Prior HCT date: __/__/__ Where performed: _____

HCT type: Auto Allo-unrelated Allo-related*Complete copies of above questions if >1 prior HCT***5. PRODUCT IDENTIFICATION**

Product/s (this course) genetically modified: Y | N

Donor type: Autologous Allo-unrelated Allo-related

If related, donor relation:

Same donor used for prior CT/HCT: Y | N | Unk | NA

Donor age:

Donor sex:

Unrelated donor: GRID:

Donor Registry:

Donor country:

Number of products: (per protocol) as part of this course of CT: ____

Complete copies of above questions if > 1 donor used

Product name: _____

Date of product request: __/__/__

Date manufacturing started: __/__/__

Final product ready for shipping: __/__/__

Final product shipped: __/__/__

Date receipt of product: __/__/__

Planned setting of infusion: Inpatient OutpatientActual setting of infusion: Inpatient Outpatient

Cell Therapy Pre-Infusion (continued)
6. PLANNED HCT

Subsequent HCT planned as part of protocol: Y | N

 Subsequent HCT type: Autologous Allogeneic

Circumstance for subsequent HCT:

- Regardless of response to cell therapy
 Only if responds to cell therapy
 Only if fails or incomplete response

7. INDICATION

Indication for cell therapy:

If Malignant disease - Complete Disease Classification Form

Date of diagnosis: __/__/__ (n/a for malignant disease)

8. BRIDGING THERAPY

Bridging therapy was given prior to CT infusion: Y | N

If yes, Date started: __/__/__

9. LYMPHODEPLETING THERAPY

Lymphodepleting therapy given prior infusion: Y | N

Drug	Total dose/mg*	Date started	Dose reduction, % and reason
		__/__/__	
		__/__/__	
		__/__/__	

* total dose = daily dose x number of days

10. TOXICITY PROPHYLAXIS

CRS prophylaxis agents given:

Neurotoxicity prophylaxis agents given:

11. LAB ASSESSMENTS PRIOR TO LYMPHODEPLETION

Date of complete blood count: __/__/__

	Value	Units
WBC		x 10 ⁹ /L
Neutrophils x 10 ⁹ /L		x 10 ⁹ /L
Lymphocytes x 10 ⁹ /L		x 10 ⁹ /L
Haemoglobin g/L		g/L
Haematocrit %		%

RBC transfused ≤ 30 days prior: Y | N

Platelets x 10 ⁹ /L		x 10 ⁹ /L
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Platelets transfused ≤ 7 days prior: Y | N | Unk

Growth factor given within 7 days prior (or long acting growth factors within 14 days): Y | N

	Value	Units	Date of sample
LDH		U/L	
LDH ULN		U/L	
Total serum ferritin		ug/L	
C-reactive protein		mg/L	
C-reactive protein ULN		mg/L	
Serum Creatinine		umol/L	

12. PATIENT ASSESSMENT

Karnofsky/Lansky Score: ____ ECOG: ____

13. COMORBID CONDITIONS

COVID-19 positive any time prior: Y | N

If yes: Hospitalised: Y | N

Mechanically ventilated: Y | N

Comorbidities (Sorrer et al)

- | | |
|--|--|
| <input type="checkbox"/> Arrhythmia | <input type="checkbox"/> Obesity |
| <input type="checkbox"/> Cardiac | <input type="checkbox"/> Peptic Ulcer |
| <input type="checkbox"/> Cerebrovascular | <input type="checkbox"/> Psychiatric |
| <input type="checkbox"/> Diabetes | <input type="checkbox"/> Pulmonary, mod |
| <input type="checkbox"/> Heart valve dis | <input type="checkbox"/> Pulmonary, severe |
| <input type="checkbox"/> Hepatic, mild | <input type="checkbox"/> Renal, mod/severe -> on dialysis: Y N |
| <input type="checkbox"/> Hepatic, mod/sev | <input type="checkbox"/> Rheumatologic |
| <input type="checkbox"/> Infection | <input type="checkbox"/> Prior malignancy, specify: _____ |
| <input type="checkbox"/> Inflammatory bowel disorder | <input type="checkbox"/> Other comorbidity: _____ |

Cell Therapy Product
1. PRODUCT SOURCE

Date product collected: ___/___/___
 Tissue source: marrow | peripheral blood | other: _____
 Cell type: Lymphocytes unselected | CD4+ | CD8+ | other: _____
 Where manufactured / processed:
 Novartis | Kite pharma | Cell processing lab on site | other: _____

2. AUTOLOGOUS PRODUCT

Method of collection: BM aspirate | Leukapheresis | other: _____
 Number of collections: _____

3. CELL MANIPULATION -

Not required for commercial products

Cells selected /modified/engineered: Y | N
 Portion manipulated: Entire product | Portion
 ⇒ If portion, unmanipulated portion also infused: Y | N
 Same manipulation method on entire/all portions of product: Y | N
 Method used: _____

Complete following if genetically manipulated:

Transfection -> Viral transduction | Non-viral transfection
 Gene editing -> specify gene
 Cells engineered to express a non-native protein: Y | N
 ->T-cell receptor | CAR,specify construct | Suicide gene, specify
 Other genetic manipulation

Manipulated to recognize specific target/antigen -> specify target: _____

4. CELL PRODUCT ANALYSIS

Not required for commercial products

Transfection efficiency performed (genetically engineered cells):
 Y | N | Unk
 If yes: Date performed: ___/___/___
 Transfection efficiency % _____ target achieved: Y | N
 Viability of cells performed: Y | N | Unk ->
 If yes: Date performed: ___/___/___
 Viability of cells % _____
 Method: 7-AAD | Propidium iodide | Trypton blue | Other

5. Out of specification

Commercial products only

Product is out of specification: Y | N | Unk
 If yes, reason: _____

6. PRODUCT INFUSION

Total number planned infusions of this product: _____
 (number of infusions specified in the protocol)

Cell Therapy Infusion
1. CELL PRODUCT IDENTIFIERS

Cell product ID _____ (e.g. Kite Konnect)
 ISBT DIN number _____
 Batch number _____ (Kymriah)
 Lot number _____ (Yescarta, Tecartus)

2. INFUSION

Date of infusion: ___/___/___
 Entire product volume infused: Y | N
 → If no, reserved portion fate:
 Discarded | Cryopreserved | other specify
 Route of infusion: IV | other specify route/site

If Product was not infused

Reason why not infused:
 Disease progression | Comorbidities | Other: _____

3. CELL DOSES

Recipient weight /kg _____
 Recipient height /cm _____
Report total number of cells given (not cells per kg)
 Total number of cells _____ x 10 ____
 Lymphocytes unselected _____ x 10 ____
 CD4+ lymphocytes _____ x 10 ____
 CD8+ lymphocytes _____ x 10 ____
 Natural killer cells (NK cells) _____ x 10 ____
 Dendritic cells / tumour cell hybridomas _____ x 10 ____
 Mesenchymal stromal stem cells (MSCs) _____ x 10 ____
 Unspecified mononuclear cells _____ x 10 ____
 Endothelial progenitor cells _____ x 10 ____
 Human umbilical cord perivascular cells _____ x 10 ____
 Cardiac progenitor cells _____ x 10 ____
 Islet cells _____ x 10 ____
 Oligodendrocytes _____ x 10 ____
 Other, specify cell type and dose _____

4. CONCOMITANT THERAPY

Recipient receive concomitant therapy: Y | N
 If yes, specify drugs: _____
 When given: Simultaneous | Post cell therapy | Unknown