ABMTRR Cell Th	nerapy Data - Pre-Infusion Page 1 of 2
Patient Information	Product: Tisagenlecleucel   Axicabtagene   other, specify
1 PATIENT IDENTIFICATION	Date of product request: / /
Hospital: UPN:	Date manufacturing started: / /
DOB: / / Sex:	Final product ready for shipping: / /
Usual residence: Postcode:	Final product shipped: / /
Bace: Indigenous status:	Date receipt of product:
Consented: Y   N	Planned setting of infusion: Innational Outpations
	A stud setting of infusion: inpatient   Outpatient
	=
Cell Therapy Pre-Infusion	6. PLANNED HCT
Referral centre:	Subsequent HCT type: Auto   Allo
Referring doctor:	Circumstance for subcoquent HCT:
Date of first referral for cell therapy: / /	
	Regardless of response to cell therapy
2. CELL THERAPY	Only if fails or incomplete response
Participating in CT clinical trial: Y   N	
If yes: Corporate   Investigator initiated   other, and details:	7. INDICATION
Study id number:	Considered as DLI: Y   N
Complete copies of above questions if on multiple trials	Indication for cell therapy:
If no, reason why not in clinical trial:	If Malignant disease - Complete Disease Form
Institutional guidelines   Hospital exemption   Compassionate use	Date of diagnosis://
Product funding: Clinical Trial   MBS   MTOP   Self-funded	8. DISEASE ASSESSMENT PRIOR TO CELL THERAPY
3. PRIOR CELL THERAPY (CT)	Bridging therapy was given prior to CT infusion: Y   N
This is first course of cell therapy (non HCT): Y   N  Unk	If yes, Date started://
If no: All reported to: ABMTRR  CIBMTR EBMT	Date Disease Considered
Number prior CTs: Date of CT://	Molocular / / VINUnkINA VIN
Where performed: Indication:	
Cell source(s): Auto   Allo-unrel   Allo-related	Flow cytometry _/_/ Y N Unk NA Y N
Complete copies of above questions if >1 prior CT	Karyotyping _/_/_ Y N Unk NA Y N
4. PRIOR TRANSPLANT (HCT)	FISH       _/_/_       Y N Unk NA       Y   N
Received prior HCT: Y   N  Unk	Radiological _/_/_ Y N Unk NA
HCT date: / / Where performed:	Clinical/haem// Y N Unk NA
HCT type: Auto   Allo-unrel   Allo-related	Disease status immediately prior CT: CR   Not in CR
Complete copies of above questions if >1 prior HCT	Date assessed://
5. PRODUCT IDENTIFICATION	9. LYMPHODEPLETING THERAPY
Product/s (this course) genetically modified: Y   N	Lymphodepleting therapy given prior infusion: Y   N
Donor type: Auto   Allo-unrel   Allo-related	
other relative   HLA m/m relative	Drug       Total       Date       Dose reduction,         dose/mg       started       % and reason
Same donor used for prior CT/HCT: Y   N  Unk  NA	
GRID:	
Donor Registry: Donor country:	
Donor age: Donor sex:	
Number of products: (per protocol) as part of this course of CT: Complete copies of above questions if > 1 donor used	

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10. PATIENT ASSESSMENT pre infusion	5. Out of specification Commercial products only
Karnofsky/Lansky Score: ECOG:	Product is out of specification: Y   N   Unk
COVID-19 positive any time prior: Y   N	If yes, reason:
Hospitalised: Y   N Mechanically ventilated: Y   N	
Comorbidities (Sorror et al) malignant indications only	Total number planned infusions of this product:
Arrhythmia Dobesity	(this course of cell therapy)
Cardiac Peptic Ulcer	(
Cerebrovascular Psychiatric	
Diabetes Delimonary, mod	
Heart valve dis Pulmonary, severe	1. CELL PRODUCT IDENTIFIERS
Hepatic, mild	Cell product ID
Hepatic, mod/sev   Prior malignancy, specify:	ISBT DIN number
Vear diagnosed _/_/_	Batch number
	Lot number
Other comorbid condition:	
	2. INFUSION
Cell Therapy Product	Date of infusion://
	Entire product volume infused: Y   N
Date product collected: / /	➔ If no, reserved portion fate:
Tissue source: BM   PB   Cord Blood   other specify	Discarded   Cryopreserved   other specify
Cell type: Lymphocytes unsel   CD4+   CD8+   TReg cells   other	Route of infusion: IV   other specify route/site
Where manufactured / processed:	Or Product was not infused
Novartis   Kite pharma   Cell processing lab on site   other	Reason why not infused:
	Disease progression   Comorbidities   Other specify
2. AUTOLOGOUS PRODUCT Method of collection: BM aspirate  Leukapheresis   other specify	
Number of collections:	3. CELL DOSES
	Recipient weight /kg
3. CELL MANIPULATION	Recipient height /cm
Cells selected /modified/engineered: Y   N	Report total number of cells given (not cells per kg)
Portion manipulated: Entire product  Portion	
$\Rightarrow$ If portion, unmanipulated portion also infused: Y   N	Lymphocytes unselected x 10
Same manipulation method on entire/all portions of product: Y $\mid$ N	CD4+ lymphocytes x 10
Method used: Cultured   Cell selection specific antigen affinity	CD8+ lymphocytes x 10
Genetic manipulation   other specify	Natural killer cells (NK cells) x 10
Transfection -> Viral transduction   Non-viral transfection	Dendritic cells / tumour cell hybridomas x 10
Gene editing -> specify gene	Mesenchymal stromal stem cells (MSCs) _ x 10
Cells engineered to express a non-native protein: Y   N	Unspecified mononuclear cells x 10
->T-cell receptor   CAR, specify construct   Suicide gene, specify	Endothelial progenitor cells v 10
Other genetic manipulation	Human umbilital card perioacular calls
Manipulated to recognize specific target/antigen -> specify target:	
	Cardiac progenitor cells x 10
4. CELL PRODUCT ANALYSIS Transfection efficiency performed (genetically engineered cells):	Islet cells x 10
Y   N  Unk	Oligodendrocytes x 10
If yes: Date performed://	Other, specify cell type and dose
Transfection efficiency % target achieved: Y   N	
Viability of cells performed: Y   N  Unk ->	4. CONCOMITANT THERAPY
II yes: Date performed:/_/ Viability of cells %	Recipient receive concomitant therapy: Y   N
Method: 7-AAD Propidium iodide! Trypton blue 10ther	ii yes, specify urugs.
	When given: Simultaneous   Post cell therapy   Unknown